



Catalog Number: AE81076SF

Size: 96 Tests

Instruction manual

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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## Dimetridazole ELISA Kit

### **【INTENDED USE】**

For the quantitative detection of **Dimetridazole** concentration in **tissue, honey, raw milk, raw egg, cooked egg**.

This package insert must be read in its entirety before using this product.

### **If You Have Problems**

Our expert Technical Support Staff is available to assist you in answering your questions and resolving issues to ensure complete customer satisfaction.

### **Please Contact Us**

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In order to obtain higher efficiency service, please ready to supply the lot number of the kit to us (found on the outside of the box).

**【REAGENTS AND MATERIALS PROVIDED】**

Reagents	Quantity	Reagents	Quantity
Assay plate (96 Wells)	1	Instruction manual	1
Standard	6 x 1 mL	Redissolving Solution	2 x 20 mL
Antibody	1 x 7 mL	HRP-Conjugate	1 x 12 mL
Wash Buffer (concentrate 20 x)	1 x 20 mL	Stop Solution	1 x 7 mL
Substrate A	1 x 7 mL	Substrate B	1 x 7 mL
Tissue sample solution I (lyophilized, Optional)	1	Milk sample solution (Optional)	1 x 10 mL
Adhesive Films	4		

**【MATERIALS REQUIRED BUT NOT SUPPLIED】**

- ※ Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at the dual-wavelength 450/630 nm.
- ※ Equipments: microplate reader, printer, homogenizer, nitrogen-drying device, vortex, centrifuge, measuring pipets, balance (a reciprocal sensibility of 0.01 g).
- ※ Micropipettors: single-channel 20-200  $\mu$ L, 100-1000  $\mu$ L, and multi-channel 250  $\mu$ L.
- ※ Reagents (AR): ethyl acetate, N-hexane, NaOH, HCl (approx 36.5%).

### **STORAGE**

<b>Unopened kit</b>	Store at 2 - 8°C. Do not use past kit expiration date.	
<b>Opened/ Reconstituted Reagents</b>	Coated assay plate	May be stored for up to 1 month at 2 - 8°C. Return unused wells to the foil pouch containing the desiccant pack, reseal along entire edge of zip-seal, and avoid the damp.
	Standard	May be stored for up to 1 month at 2 - 8°C.
	Redissolving Solution	
	Antibody	
	HRP-Conjugate	
	Wash Buffer	
	Substrate A	
	Substrate B	
	Stop Solution	
	Tissue sample solution I	
Milk sample solution		

### **INTRODUCTION**

Dimetridazole is a nitroimidazole class drug that combats protozoan infections. Target: Antiparasitic Dimetridazole (DMZ) is a 5-nitroimidazole drug traditionally used for the prevention and treatment of histomoniasis in turkeys, trichomoniasis in pigeons, genital trichomoniasis in cattle and the prevention and treatment of

haemorrhagic enteritis in pigs.

### **【PRINCIPLE OF THE ASSAY】**

This assay is based on the competitive enzyme immunoassay for the detection of Dimetridazole in the sample. The coupling antigens are pre-coated on the micro-well stripes. The Dimetridazole in the sample and the coupling antigens pre-coated on the micro-well stripes compete for the anti-Dimetridazole antibodies. After the addition of the enzyme conjugate, the TMB substrate is added for coloration. The optical density (OD) value of the sample has a negative correlation with the Dimetridazole in it. This value is compared to the standard curve and the Dimetridazole concentration is subsequently obtained.

### **【LIMITATIONS OF THE PROCEDURE】**

- ※ FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- ※ The kit should not be used beyond the expiration date on the kit label.
- ※ Do not mix or substitute reagents with those from other lots or sources.
- ※ It is important that the Calibrator Diluent selected for the standard curve be consistent with the samples being assayed.
- ※ If samples generate values higher than the highest standard, dilute the samples with the appropriate Calibrator Diluent and repeat the assay.
- ※ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- ※ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors have been tested in the ELISA Kit, the possibility of interference cannot be excluded.

### **【DETECTION RANGE】**

0.01 ppb - 0.81 ppb. The standard curve concentrations used for the ELISA's were 0.81 ppb, 0.27 ppb, 0.09 ppb, 0.03 ppb, 0.01 ppb, 0 ppb.

Tube	S5	S4	S3	S2	S1	S0
ppb	0.81	0.27	0.09	0.03	0.01	0

**【 TECHNICAL SPECIFICATIONS 】**

**Sensitivity:** 0.01 ppb

**Detection limit:**

Tissue	0.1 ppb
Honey	0.1 ppb
Raw milk (Method 1)	0.2 ppb
Raw milk (Method 2)	0.05 ppb
Raw egg (Method 1)	0.1 ppb
Raw egg (Method 2)	0.2 ppb
Cooked egg	0.1 ppb

**Cross-reaction rate:**

Dimetridazole	100%
Metronidazole	10%
Hydroxy Metronidazol	4.7%
HMMNI	5.3%
Tinidazole	0.2%
Ornidazole	0.5%

**Recovery rate:**

Tissue	90±25%
Honey	90±30%
Raw milk	85±20%
Raw egg	90±15%
Cooked egg	90±25%

**【PRECISION】****Intra-assay Precision** (Precision within an assay)

Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

**Inter-assay Precision** (Precision between assays)

Three samples of known concentration were tested in forty separate assays to assess inter-assay precision.

$$CV(\%) = SD/\text{mean} \times 100$$

**Intra-Assay: CV<8%**

**Inter-Assay: CV<15%**

**【STABILITY】**

The stability of ELISA kit is determined by the loss rate of activity. The loss rate of this kit is less than 5% within the expiration date under appropriate storage condition.

The loss rate was determined by accelerated thermal degradation test. Keep the kit at 37°C for 4 and 7 days, and compare O.D. values of the kit kept at 37°C with that of at recommended temperature. (referring from China Biological Products Standard, which was calculated by the Arrhenius equation. For ELISA kit, 4 days storage at 37°C can be considered as 6 months at 2 - 8°C, which means 7 days at 37°C equaling 12 months at 2 - 8°C).

**Note:**

To minimize extra influence on the performance, operation procedures and lab

conditions, especially room temperature, air humidity, incubator temperature should be strictly controlled. It is also strongly suggested that the whole assay is performed by the same operator from the beginning to the end.

### **[SAMPLE PRE-TREATMENT]**

**Instructions** (The following points must be dealt with before the pre-treatment of any kind of sample):

1. Only the disposable tips can be used for the experiments and the tips must be changed when used for absorbing different reagents;
2. Before the experiment, each experimental utensil must be clean and should be re-cleaned if necessary, in order to avoid the contamination that interferes with the experimental results.

#### **Solution preparation before sample pre-treatment:**

1. 1 M HCl solution: dissolve 1 mL HCl (36.5%) in deionized water to 11 mL.
2. 0.1 M NaOH solution: dissolve 0.8 g NaOH in deionized water to 200 mL.
3. Tissue sample solution I: dissolve the powder in 200 mL deionized water
4. Tissue sample solution II: dissolve 400 mL ethyl acetate and 100 mL N-hexane, mix.
5. Washing buffer: dilute 20 mL of the concentrated washing buffer (20×concentrated) with the distilled or deionized water at 1:19 to 400 mL (or just to the required volume) for use.

#### **Samples preparation:**

**Tissue** (chicken, pork, etc) ;

1. Homogenize the samples.
2. Weigh  $2.0 \pm 0.05$  g of the homogenized samples into 50 mL centrifugal tube.
3. Add 2 mL of the Tissue sample solution I, shake properly for 1 minute, and then add 8 mL of the Tissue sample solution II, shake properly for 3 minutes, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
4. Take 4 mL of the supernatant into a new centrifugal tube, and evaporate to dryness by nitrogen or air at 50-60°C waterbath.
5. Dissolve the dry residues in 2 mL N-hexane, add 0.5 mL of the

redissolving solution, shake properly for 1 minute, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.

6. Discard the whole upper layers, take 50  $\mu$ L of the lower layer for further analysis.

***Fold of dilution of the sample: 0.5***

**Honey;**

1. Take  $2 \pm 0.05$ g honey into 50 mL centrifugal tube, add 2 mL 0.1 M NaOH solution, shake properly for 5 minutes, dissolve it completely.
2. Add 4 mL ethyl acetate, shake properly for 5 minutes and centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
3. Take 2 mL of the supernatant into a new centrifugal tube, and evaporate to dryness by nitrogen or air at 50-60°C waterbath .
4. Dissolve the dry residues in 0.5 mL of the redissolving solution, shake properly for 2 minutes.
5. Discard the whole upper layers, take 50  $\mu$ L of the lower layer for further analysis.

***Fold of dilution of the sample: 0.5***

**Raw milk (Method 1);**

1. Defrost the sampled raw milk samples, Let stand to the room temperature (20-25°C) for at least 30 minutes.
2. Take 1 mL of the lower layer sample into 2 mL centrifugal tube, add 50  $\mu$ L 1 M HCl solution, shake properly for 1 minute, centrifuge at above 4000 r/min at room temperature (20-25°C) for 10 minutes.
3. Transfer 100  $\mu$ L of the supernatant into 2 mL centrifuge tube. Add 400  $\mu$ L of the redissolving solution, shake strongly for 1 minute, dissolve it completely.
4. Take 50  $\mu$ L for further analysis.

***Fold of dilution of the sample: 5***

**Raw milk (Method 2);**

1. Defrost the sampled raw milk samples, Let stand to the room temperature (20-25°C) for at least 30 minutes.
2. Take 2 mL of the lower layer sample into 50 mL centrifugal tube, add 100



- $\mu\text{L}$  Milk sample solution and 6 mL ethyl acetate, shake properly for 5 minutes, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
3. Take 1.5 mL of the supernatant into a new centrifugal tube, and evaporate to dryness by nitrogen or air at 50-60°C waterbath.
  4. Dissolve the dry residues in 1 mL N-hexane, add 0.5 mL of the redissolving solution, shake properly for 1 minute, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
  5. Discard the whole upper layers, take 50  $\mu\text{L}$  of the lower layer for further analysis.

***Fold of dilution of the sample: 1***

**Raw egg (Method 1);**

1. Homogenize the sample (Egg white, egg yolk or whole egg).
2. Take 1 mL of the homogenized samples into 50 mL centrifugal tube, add 1 mL 0.1 M NaOH solution and 8 mL ethyl acetate, shake properly for 5 minutes, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
3. Take 4 mL of the supernatant into a new centrifugal tube, and evaporate to dryness by nitrogen or air at 50-60°C waterbath.
4. Dissolve the dry residues in 1 mL N-hexane, add 0.5 mL of the redissolving solution, shake properly for 2 minutes, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
5. Discard the whole upper layers, take 50  $\mu\text{L}$  of the lower layer for further analysis.

***Fold of dilution of the sample: 1***

**Raw egg (Method 2);**

1. Homogenize the sample (Egg white, egg yolk or whole egg).
2. Take 1 mL of the homogenized samples into 50 mL centrifugal tube, add 100  $\mu\text{L}$  1 M HCl solution, shake properly for 1 minute, centrifuge at above 4000 r/min at room temperature (20-25°C) for 10 minutes.
3. Take 100  $\mu\text{L}$  of the supernatant into a new centrifugal tube, add 400  $\mu\text{L}$  of the redissolving solution, shake properly for 1 minute.
4. Take 50  $\mu\text{L}$  for further analysis.

***Fold of dilution of the sample: 1***

**Cooked egg;**

1. Homogenize the sample.
2. Weigh  $1.0 \pm 0.05$  g of the homogenized sample into 50 mL centrifugal tube, add 1 mL 0.1 M NaOH solution and 8 mL ethyl acetate, shake properly for 5 minutes, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
3. Take 2 mL of the supernatant into a new centrifugal tube, and evaporate to dryness by nitrogen or air at 50-60°C waterbath.
4. Dissolve the dry residues in 1 mL N-hexane, add 0.5 mL of the redissolving solution, shake properly for 2 minutes, centrifuge at above 3000 r/min at room temperature (20-25°C) for 5 minutes.
5. Discard the whole upper layers, take 50  $\mu$ L of the lower layer for further analysis.

***Fold of dilution of the sample: 2***

**[SAMPLE STORAGE]**

1. Untreated samples are stored at frozen environment.
2. Prepared sample can be stable at 2-8°C for 1 week.

**[TABLE]**

**Table depicting an example of the arrangement of blanks, standards and samples in the microwell strips:**

	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>
<b>A</b>	Standard 5 (0.81 ppb)	Standard 5 (0.81 ppb)	Sample 3	Sample 3
<b>B</b>	Standard 4 (0.27 ppb)	Standard 4 (0.27 ppb)	Sample 4	Sample 4
<b>C</b>	Standard 3 (0.09 ppb)	Standard 3 (0.09 ppb)	Sample 5	Sample 5
<b>D</b>	Standard 2 (0.03 ppb)	Standard 2 (0.03 ppb)	Sample 6	Sample 6
<b>E</b>	Standard 1	Standard 1	Sample 7	Sample 7

	(0.01 ppb)	(0.01 ppb)		
<b>F</b>	Standard 0 (0 ppb)	Standard 0 (0 ppb)	Sample 8	Sample 8
<b>G</b>	Sample 1	Sample 1	Sample 9	Sample 9
<b>H</b>	Sample 2	Sample 2	Sample 10	Sample 10

### **【ASSAY PROCEDURE】**

**Bring all reagents and samples to room temperature before use. It is recommended that all samples, controls, and standards be assayed in duplicate.**

1. Bring test kit to the room temperature (20-25°C) for at least 30 minutes, note that each reagent must be shaken evenly before use; put the required micro-well strips into plate frames. Re-sealed the unused microplate, stored at 2-8°C, not frozen.
2. Add 50 µL of the sample or standard solution to separate duplicate wells, and add 50 µL of the antibody working solution into each well. Mix by shaking gently, seal the microplate with the cover membrane, and incubate at 4°C for 1 hour.
3. Pour the liquid, wash the microplate with the washing buffer at 250 µL/well for 3-4 times. Each time soak the well with the washing buffer for 15-30s, flap to dry with absorbent paper (if there are the bubbles after flapping, cut them with the clean tips).
4. Add 100 µL of HRP-Conjugate to each well. seal the microplate with the cover membrane. Incubate for 20 minutes at 25°C.
5. Pour the liquid, wash the microplate with the washing buffer at 250 µL/well for 5 times. Each time soak the well with the washing buffer for 15-30s, flap to dry with absorbent paper (if there are the bubbles after flapping, cut them with the clean tips).
6. Add 50 µL of Substrate A and 50 µL of Substrate B to each well, mix well. Incubate for 15 minutes at 25°C. Keeping the plate away from drafts and other temperature fluctuations in the dark. Avoid placing the plate in direct light.
7. Add 50 µL of Stop Solution to each well. when the first four wells containing the highest concentration of standards develop obvious blue

color. If color change does not appear uniform, gently tap the plate to ensure thorough mixing.

- Determine the optical density of each well within 5 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 630 nm. Subtract readings at 630 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

### **CALCULATION OF RESULTS**

**There are two methods to judge the results; the first one is the rough judgment, while the second is the quantitative determination. Note that the OD value of the sample has a negative correlation with the Dimetridazole concentration in the sample.**

#### **1) Qualitative determination**

The concentration range (ppb) of Dimetridazole can be obtained from comparing the average OD value of the sample with that of the standard solution.

#### **2) Quantitative determination**

The mean values of the absorbance values obtained for the average OD value (B) of the sample and the standard solution divided by the OD value (B0) of the first standard solution (0 standard) and subsequently multiplied by 100%, that is

Percentage of absorbance value =	B	×100%
	B0	

B—the average (double wells) OD value of the sample or the standard solution

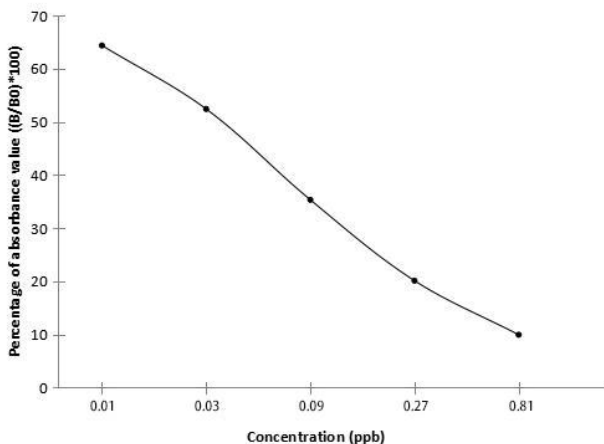
B0—the average OD value of the 0 ppb standard solution

Draw the standard curve with the absorption percentages of standard solutions and the semilogarithm values of Dimetridazole standard solutions (ppb) as Y- and X-axis, respectively. Read the corresponding concentration of the sample from the standard curve by incorporating its absorption percentage into the standard curve. The resulting value is subsequently multiplied by the dilution fold, finally obtaining Dimetridazole concentration in the sample.

Using the professional software of this kit will be more convenient for accurate and rapid analysis of a large amount of samples.

### **TYPICAL DATA**

For convenience in result calculation, absorbance as abscissa and standard concentrations can be used as ordinate. The standard curve provided in the manual is only for reference, experimenters should draw the standard curve based on data of themselves.



**Precautions:** This kit contains trace amount of proclin® 300 which is toxic and direct touch of reagents by mouth, eyes, and skin should be avoided. If get touched or get into the mouth, please rinse with plenty of water. The Stop Buffer is a corrosive sulfuric acid solution, please take care for safety.

### **IMPORTANT NOTE**

1. The instruction manual also suits for the kit of 48T, but all reagents of 48T kit are reduced by half.
2. There may be some foggy substance in the wells when the plate is opened at the first time. It will not effect on the final assay results. Do not

- remove microtiter plate from the storage bag unless needed.
3. Do not mix or substitute reagents from one kit lot to another. Use only the reagents supplied by manufacturer.
  4. Samples or reagents addition: Please use the freshly prepared Standard. Please carefully add samples to wells and mix gently to avoid foaming. Do not touch the well wall as far as possible. For each step in the procedure, total dispensing time for addition of reagents or samples to the assay plate should not exceed 10 minutes. This will ensure equal elapsed time for each pipetting step, without interruption. Duplication of all standards and specimens, although not required, is recommended. To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
  5. Incubation: To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods between incubation steps. Once reagents have been added to the well strips, DO NOT let the strips DRY at any time during the assay. Incubation time and temperature must be observed.
  6. Washing: The wash procedure is critical. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Solution by aspirating or decanting and remove any drop of water and fingerprint on the bottom of the plate. Insufficient washing will result in poor precision and falsely elevated absorbance reading. When using an automated plate washer, adding a 30 second soak period following the addition of wash buffer, and/or rotating the plate 180 degrees between wash steps may improve assay precision.
  7. Controlling of reaction time: Observe the change of color after adding Substrate Solution (e.g. observation once every 10 minutes), Substrate Solution should change from colorless or light blue to gradations of blue. If the color is too deep, add Stop Solution in advance to avoid excessively strong reaction which will result in inaccurate absorbance reading.
  8. Substrate Solution is easily contaminated. Substrate Solution should remain colorless or light blue until added to the plate. Please protect it from light.

9. Stop Solution should be added to the plate in the same order as the Substrate Solution. The color developed in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that are green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.
10. Protect all reagents from strong light during storage and incubation. All the bottle caps of reagents should be covered tightly to prevent the evaporation and contamination of microorganism.
11. Wrong operations during the reagents preparation and loading, as well as incorrect parameter setting for the plate reader may lead to incorrect results. A microplate plate reader with a bandwidth of 10nm or less and an optical density range of 0-3 O.D. or greater at  $450 \pm 10\text{nm}$  wavelength is acceptable for use in absorbance measurement.
12. Even the same operator might get different results in two separate experiments. In order to get better reproducible results, the operation of every step in the assay should be controlled. Furthermore, a preliminary experiment before assay for each batch is recommended.
13. Limited by the current condition and scientific technology, we can't completely conduct the comprehensive identification and analysis on the raw material provided by suppliers. So there might be some qualitative and technical risks to use the kit.
14. The final experimental results will be closely related to validity of the products, operation skills of the end users and the experimental environments. Please make sure that sufficient samples are available.
15. Kits from different batches may be a little different in detection range, sensitivity and color developing time.
16. Each kit has been strictly passed Q.C test. However, results from end users might be inconsistent with our in-house data due to some unexpected transportation conditions or different lab equipments. Intra-assay variance among kits from different batches might arise from above factors, too.
17. Kits from different manufacturers with the same item might produce different results, since we haven't compared our products with other manufacturers.

18. The Stop Solution provided with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material.
19. Valid period: six months.





